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The toxicity of lithium to human cardiomyocytes

Junwei Shen^{1,2†}, Xin Li^{3†}, Xiujuan Shi¹, Wei Wang¹, Hua Zhou³, Jiawen Wu¹, Xin Wang³ and Jue Li^{1,2*}

Abstract

Background: Lithium is widely used in the electronic consumer market and electric vehicles and has a great contribution to the world economy, resulting in large quantities of lithium waste in the environment. The Yangtze River Basin is one of the most developed areas in China. However, the environmental influence of lithium in the Yangtze River Basin and its roles in cardiomyocytes have not yet been clarified.

Results: Here, we found that the concentration of lithium in the water environment is very high in Shanghai, as well as in tap water, which might be caused by the pollution of lithium batteries. Lithium inhibits cell viability and proliferation of human cardiomyocytes. Moreover, lithium promotes cell apoptosis significantly. And we found that lithium controls cardiomyocytes' functions through regulating glycogen synthase kinase 3 beta signaling.

Conclusions: This study reveals that the water environment of Shanghai might be polluted by the lithium batteries; and the enrichment of lithium might cause damage to human cardiomyocytes. It is imperative to detect lithium concentration in the water environments (such as tap water and irrigation water) and effectively recycle lithium batteries in the future.

Keywords: Lithium, Pollution, The Yangtze River Basin, Cardiomyocyte

Background

Lithium is the first element in the alkali metal group and the lightest metal. It is very active; therefore, it exists mainly in the form of compounds (such as apatite or aluminum silicide) in the environment [1–4]. The other two elements in the alkali metal group, sodium and potassium, account for more than 2% in the earth's crust; however, in contrast to them, the existence of lithium on the earth is scarce, counting only 0.0065% [5, 6]. Although lithium may also be enriched in several mines and salt lakes, it is usually widely spread in trace amounts in rocks and soils, as well as in water, including surface water, groundwater, and seawater [7]. For example, the content

in seawater is very low, only 0.17~mg/L. In freshwater, the content is scarce, less than 0.04~mg/L [7]. The lithium compounds are highly soluble; therefore the main form of lithium in water is ion [8]. Since the concentration of lithium in the natural environment is generally very low, it does not cause damage to the environment.

However, since the first commercial lithium battery was invented in 1991, it has rapidly become popular and is widely used in consumer electronics such as laptop computers, smartphones, and tablet computers [9, 10]. The large-scale application of lithium batteries makes a great contribution to the world economy [11]. However, it might cause lots of lithium pollution. In cities, lithium batteries are routinely discarded in the environment, along with other solid garbage [10]. The lithium batteries are still cheap enough to throw old batteries and obtain virgin material [10]. Therefore, the recycling rate is meager, even in developed countries that do well in environmental protection [10]. The rapid increase in consumption and the serious shortage of recycling might

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lead to continuous accumulation of lithium in the developed areas where lithium is broadly used.

As one of the most economically developed regions in China, the Yangtze River Basin has a GDP accounting for two-fifths of China [12]. It contains a series of megacities, including Shanghai, Suzhou, Changsha, Nanjing, Wuhan and Chongqing. This region has a well-developed electronic consumer market and rapid economic growth, which continues to promote lithium consumption. However, the research on the impact of lithium pollution in the Yangtze River Basin has not yet been reported to date. Shanghai, at the estuary of the Yangtze River, is the leader of the Chinese economy. Shanghai is one of the biggest consumer electronics market in China. Besides, Shanghai is actively promoting the popularization of electric vehicles these years [13]. The facts above increase substantially the use of lithium batteries, which accumulates the lithium pollution in this area. Due to the discharge of large quantities of lithium resources, lithium pollution is growing rapidly and imposing a severe threat to the environment and humans [14].

Though the low concentration of lithium has no harmful effect on the environment, at higher levels lithium may cause considerable damage to the aquatic and terrestrial environment [15]. For example, a small dose of lithium has a significant inhibitory effect on the proliferation and growth of aquatic organisms including Pimephales promelas, Ceriodaphnia dubia, and Elimia clavaeformis [16]. Also, lithium in water can accumulate in plants and cause damage to plant growth and development [17, 18]. For example, 60 mmol/L of lithium can damage the growth of sunflower; the same concentration can also affect the growth of corn [17]. Lithium can be enriched in animals by food chains, and high concentrations of lithium can also cause severe damage to animals [19]. For instance, rats were treated with small doses of lithium for 7 weeks (every alternate day) [20]. The epithelium lining of renal tissue was injured, and some significant changes were observed in the glomerular region in the corticomedullary region [20]. Besides, high concentrations of lithium could cause severe damage to humans, including the nervous system (including coarse tremor and hyperreflexia), kidney (including sodium-losing nephritis and nephrotic syndrome), and endocrine system (including hypothyroidism) [21-24]. However, the effects of lithium on the cardiovascular system have not been studied yet.

As one of the most important organs in our body, the heart is the first organ that functions in the embryo development. It is exposed to an open environment that contains various types of harmful factors, such as nicotine, alcohol and drugs. Notably, the cardiomyocytes that contracts in the heart, lacks regeneration capability [25]. Therefore, each damage to the cardiomyocytes could accumulate constantly during the lifetime. Moreover, the damage to cardiac contractility might be irreversible. Therefore, cardiotoxicity becomes the most important cause for the recall of prescription drugs [26]. It accounts approximately half of all drugs withdrawn in the last two decades [26]. For example, the obesity treatment drug sibutramine and the antidiabetic drug rosiglitazone were both recalled due to their cardiotoxicity effects in 2010 [27, 28]. However, the effects of lithium on cardiomyocytes remain to be determined. Our research detected that lithium concentrations in the Yangtze River and rivers in Shanghai are relatively high, and it is possible to cause harm to human health through food chains [29]. Then we found that lithium not only significantly inhibited cell viability and cell proliferation of human cardiomyocytes, but also promoted cell apoptosis. Finally, we found that these effects of lithium may be related to the regulation of GSK3β.

Materials and methods

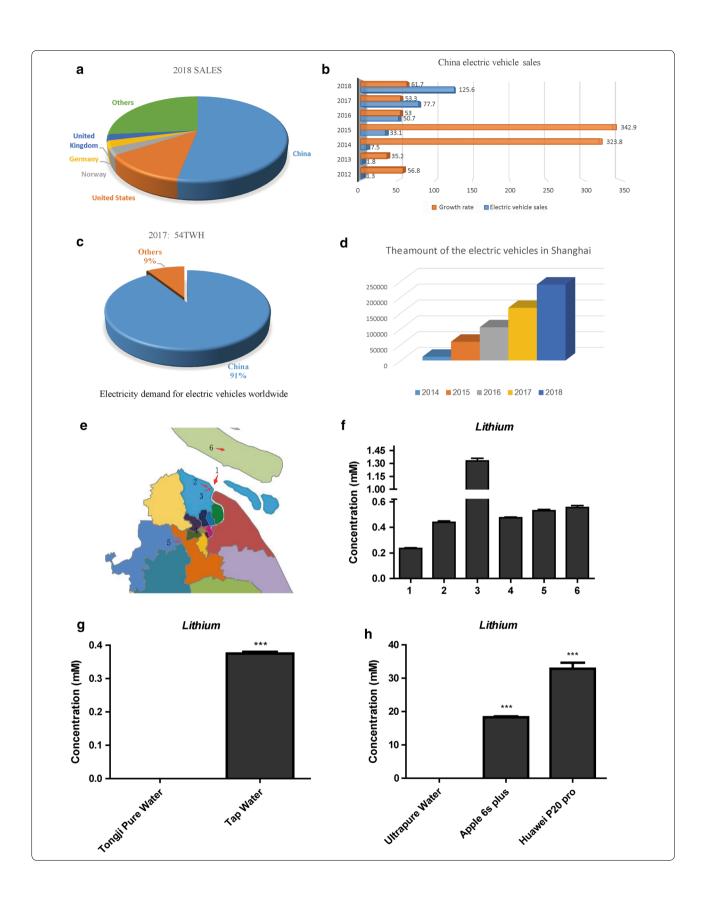
Detection of the concentration of lithium in water

The water samples were obtained from the places shown in Fig. 1e. The concentration of lithium in water samples was detected with K-Lite8F (Cornley, Meizhou, China) according to the manual. To evaluate the effects of lithium batteries on water, two disabled Apple 6s plus batteries and two disabled Huawei P20 pro batteries were used in this study. Each battery was placed in a beaker, soaked in ultrapure water, and then the battery is punctured with a clean needle, then added ultrapure water to 2 L. After 3 days, the polluted water was filtered by filter paper. The concentration of lithium in these samples was detected with K-Lite8F (Cornley) as well.

(See figure on next page.)

Fig. 1 Lithium enrichment in Shanghai water environment. **a** Pie chart shows the sales of electric vehicle worldwide in 2018. **b** Histogram shows the China electric vehicle sales and growth rate from 2012 to 2018. **c** Pie chart shows the demand for electric vehicles worldwide in 2017. **d** Histogram shows the electric vehicles in Shanghai from 2014 to 2018. **e** Schematic diagram shows the six water environments. (1) Yangtze river, (2) freshwater lake, (3) pit lake, (4) Suzhou River, (5) Dingpu River, (6) Chongming island. **f** Histogram shows the concentration of lithium in the six water environments. **g** Histogram shows the concentration of lithium in Tongji pure water and tap water. **h** Histogram shows the concentration of lithium in ultrapure water, Apple 6s plus and Huawei P20 pro

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Cell culture and treatment

Human AC16 cardiomyocyte line was cultured in high-glucose Dulbecco's modified Eagle's medium (Gibco, Grand Island, NY) supplemented with 10% (v/v) fetal bovine serum (Gibco) and 1% (v/v) penicillin and streptomycin (Gibco). Cells were grown in a humidified atmosphere of 5% $\rm CO_2$ at 37 °C. LiCl (Sinopharm Chemical Reagent, Shanghai, China) or $\rm Li_2SO_4$ (Sinopharm Chemical Reagent, Shanghai, China) was reconstituted in ddH₂O. AC16 cells were incubated with LiCl or $\rm Li_2SO_4$ at different concentrations (0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L) for 48 h as specified in the figure legends.

Cell viability assay

The cell viability was tested with CellTiter-LumiTM Luminescent Cell Viability Assay Kit (Beyotime, Nantong, China). The AC16 cells were seeded into 96-well plates $(1\times10^3~\text{cells/well})$ [30]. After 48 h treatment with control (ddH₂O), LiCl, NaCl or Li₂SO₄ at different concentrations (0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L), the cells were assessed. 100 μ L CellTiter-LumiTM reagent was added into each well of the plate. Then the plate was incubated at 37 °C for 10 min. The luminometer was subsequently recorded with SpectraMax M5 plate reader (Molecular Devices).

The cell viability was measured via the Cell Counting Kit-8 (CCK-8) assay as well. AC16 cells were seeded into 96-well plates (1 \times 10 3 cells/well) [31]. After 48 h treatment with control, LiCl, NaCl or Li $_2$ SO $_4$ at different concentrations (0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L), the cells were assessed. 10 μL CCK-8 reagent (Dojindo, Kumamoto, Japan) was added into each well of the plate. Then the plate was incubated at 37 $^{\circ}$ C for 2 h. The absorbance at 450 nm was subsequently recorded with SpectraMax M5 plate reader (Molecular Devices).

Cell apoptosis assay

AC16 cells were treated with control, 5 mmol/L LiCl or 2.5 mmol/L Li $_2$ SO $_4$ for 48 h. To quantify the cell apoptotic degree, the harvested cells were stained with annexin V-FITC/PI Cell Apoptosis Kit (Keygen, Nanjing, China) according to the manufacturer's instructions. After incubation for 30 min at 4 °C, the cells were analyzed using FCM (FACS Canto; BD Biosciences).

EdU proliferation assay

5-Ethynyl-2'-deoxyuridine (EdU) is a synthetic thymidine analog that can incorporate into newly synthesized DNA during S phase. Therefore, EdU detection can be used for tracking DNA replication directly [32]. The

immunofluorescence staining of EdU was performed with BeyoClick[™] EdU-555 proliferation kit (Beyotime) followed the Kit manual. Briefly, cells were cultured in 24-well plates, fixed in 4% paraformaldehyde after 48 h treatment with control, 5 mmol/L LiCl or 2.5 mmol/L Li $_2$ SO $_4$, and permeabilized with 0.2% Triton X-100 for 15 min. Cells were counterstained with Hoechst for 5 min, then were washed with and imaged in PBS. At last, the images were taken using fluorescence microscopy (Nikon).

Western blotting

Cultured AC16 cells were lysed in strong RIPA buffer containing Halt Protease Inhibitor Cocktails (Thermo, Waltham, MA). Protein concentrations were measured using a BCA protein assay kit (Pierce, Rockford, IL). Primary antibodies targeting proliferating cell nuclear antigen (PCNA) (ab29, abcam, Cambridge, UK), tumor protein p53 (TP53) (ab1101, abcam), CYCLIN E (ab33911, abcam), glycogen synthase kinase 3 beta (Gsk3β) (ab32391, abcam), GSK3β (P-Ser9) (ab131097, abcam) and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (ab9485, abcam) were incubated with proteins overnight at 4 °C, followed by incubation with the appropriate HRP (horseradish peroxidase) conjugated secondary antibodies (BA1038 and BA1039, Boster, Wuhan, China). Detection of HRP was performed using the ECL assay kit (Beyotime) and an ImageQuant LAS 4000 mini (GE, Boston, Unite States).

Statistical analysis

GraphPad Prism 7.0 software was used for statistical analysis. Statistical significance between two groups was determined using an unpaired two-tailed Student's t test. Data are presented as mean \pm SD (standard deviation) or mean \pm SEM (standard error of the mean) as indicated in the figure legends. p values were considered statistically significant if p < 0.05.

Results

The enrichment of lithium in Shanghai water environment

To investigate the potential pollution of lithium in China, we evaluated the electric vehicle market in 2018. Surprisingly, China accounts for more than half of the global market (Fig. 1a). At the same time, the electric vehicle market in China is growing rapidly these years (Fig. 1b). China accounts for 91% of the world's lithium battery demand in 2017 due to the huge electric vehicles, electric bikes and electric buses (Fig. 1c). Shanghai is one of the most active cities to promote the development of electric vehicles. The number of electric vehicles was more than 100,000 in 2016 in Shanghai, making it the largest electric vehicle city that time (Fig. 1d).

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The previous results suggest that Shanghai could become a region with lithium pollution, therefore we detected the lithium concentration in different water environments (Fig. 1e, f). Though the concentration of lithium in Yangtze River (1.80 mg/L) was the lowest in the six environments (Fig. 1e, f), it was much higher than the normal concentration in freshwater (0.04 mg/L) [7]. The concentrations of lithium in other water environments were much higher than in the Yangtze River (Fig. 1e, f), and concentration of lithium in the pit lake was the highest. Intriguingly, compared with lithium in pure water (0 mmol/L), the lithium concentration in tap water was a little high (0.37 mmol/L). Lithium concentration in the water that was contaminated by lithium batteries was very high (Fig. 1g). These data indicated that the water environment in Shanghai could be polluted by lithium batteries.

Lithium inhibits the cell viability of human cardiomyocytes

To investigate the effects of lithium on cardiomyocytes, we evaluated the cell viability of AC16 cells under LiCl exposure at different concentrations (0 mmol/L, 0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L). Compared to the control (0 mmol/L), the growth of AC16 cells was significantly inhibited by LiCl at concentrations of 5 mmol/L and 25 mmol/L with the luminescent assay, as well as the CCK-8 assay (Fig. 2a-c). To evaluate whether Cl⁻ causes the inhibitory effects, NaCl was added in the study. Interestingly, high dose of NaCl had no significant effect on cell viability of AC16 cells (Fig. 2d-f). Additionally, we observed that Li₂SO₄ at concentrations of 2.5 mmol/L and 12.5 mmol/L also obviously inhibited the cell viability of AC16 cells (Fig. 2g-i). Thus, these data suggested that lithium may exert adverse effects on cardiomyocytes to inhibit cell viability.

Lithium inhibits the cell proliferation of human cardiomyocytes

EdU proliferation assay showed that the EdU level in the 5 mmol/L LiCl treated group was significantly reduced, suggesting that the cell proliferation rate of AC16 cells reduce sharply (Fig. 3a). Similarly, 2.5 mmol/L Li₂SO₄ also inhibited AC16 cell proliferation dramatically (Fig. 3b). Then we evaluated the cell proliferation ability by cell counting. Consistently, the lithium repressed the AC16 cell proliferation significantly (Fig. 3c, d). Proliferating cell nuclear antigen (PCNA) is an essential cofactor for DNA polymerases during replication [33]. We showed that the protein expression of PCNA in AC16 cells reduced significantly after 48 h treatment with LiCl or Li₂SO₄ compared to the control (Fig. 3e, f).

Lithium triggers the cell apoptosis of human cardiomyocytes

To further evaluate the effects of lithium on cardiomyocytes, we test the cell apoptosis with Annexin V-FITC/PI apoptosis assay. AC16 cell apoptosis increased significantly after 48 h treatment with 5 mmol/L LiCl compared to the control (Fig. 4a, b). Consistently, we observed that 2.5 mmol/L Li₂SO₄ also induced AC16 cell apoptosis (Fig. 4c, d). These data suggest that lithium might promote the cell apoptosis of human cardiomyocytes.

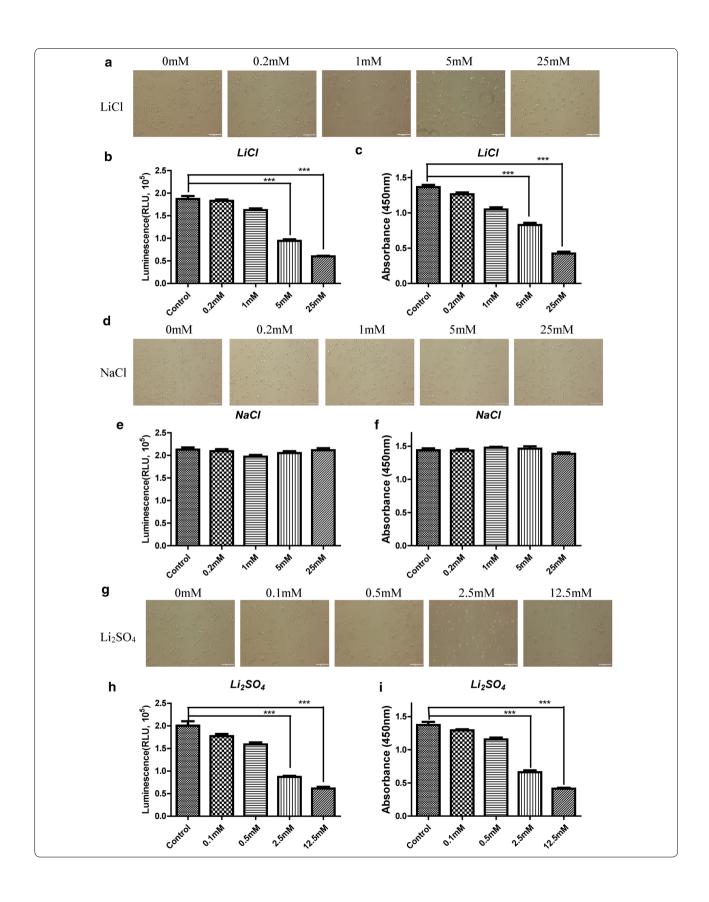
Lithium controls human cardiomyocytes via $Gsk3\beta$ signaling

TP53 is an apoptosis marker. Western blot analysis revealed that 5 mmol/L LiCl or Li_2SO_4 treatment for 48 h significantly increased the expression levels of TP53 in AC16 cells (Fig. 5a, b). Cyclin E is a proliferation marker. Cyclin E protein levels decreased after 5 mmol/L LiCl or Li_2SO_4 treatment for 48 h compared with the control (Fig. 5c, d). It is reported that glycogen synthase kinase 3 beta (Gsk3β) is active regulator of cell proliferation and

(See figure on next page.)

Fig. 2 Lithium suppresses the cell viability of human cardiomyocytes. a Microscopic images of AC16 cells treated with control (0 mmol/L) and LiCl at different concentrations (0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L) for 48 h. Bars = 200 µm. b Influence of LiCl at different concentrations (0 mmol/L, 0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L) on cell viability of AC16 cells as measured by luminescence assay 48 h post-cell seeding. The data are represented as mean \pm SEM (n=3). ***p < 0.001. \mathbf{c} Influence of LiCl at different concentrations (0 mmol/L, 0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L) on cell viability of AC16 cells as measured by CCK-8 assay 48 h post-cell seeding. The data are represented as mean ± SEM (n = 3). ***p < 0.001. d Microscopic images of AC16 cells treated with control (0 mmol/L) and NaCl at different concentrations (0.2 mmol/L) 1 mmol/L, 5 mmol/L or 25 mmol/L) for 48 h. Bars = 200 μ m. **e** Influence of NaCl at different concentrations (0 mmol/L, 0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L) on cell viability of AC16 cells as measured by luminescence assay 48 h post-cell seeding. The data are represented as mean ± SEM (n = 3), ***p < 0.001. f Influence of NaCl at different concentrations (0 mmol/L, 0.2 mmol/L, 1 mmol/L, 5 mmol/L or 25 mmol/L) on cell viability of AC16 cells as measured by CCK-8 assay 48 h post-cell seeding. The data are represented as mean \pm SEM (n = 3). *p < 0.05, **p < 0.05, **p < 0.01, ***p < 0.001. **g** Microscopic images of AC16 cells treated with control (0 mmol/L) and Li₂SO₄ at different concentrations (0.1 mmol/L, 0.5 mmol/L, 2.5 mmol/L or 12.5 mmol/L) for 48 h. Bars = 200 μ m. **h** Influence of Li₂SO₄ at different concentrations (0 mmol/L, 0.1 mmol/L, 0.5 mmol/L, 2.5 mmol/L or 12.5 mmol/L) on cell viability of AC16 cells as measured by luminescence assay 48 h post-cell seeding. The data are represented as mean \pm SEM (n = 3). ***p < 0.001. i Influence of Li₂SO₄ at different concentrations (0 mmol/L, 0.1 mmol/L, 0.5 mmol/L, 2.5 mmol/L or 12.5 mmol/L) on cell viability of AC16 cells as measured by CCK-8 assay 48 h post-cell seeding. The data are represented as mean \pm SEM (n = 3). *p < 0.05, **p < 0.01, ***p < 0.001

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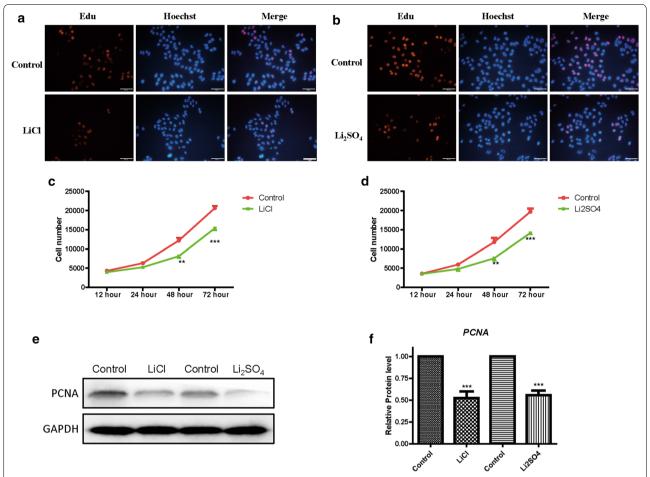


Fig. 3 Lithium suppresses the cell proliferation of human cardiomyocytes. **a** EdU detection in AC16 cells treated with control and 5 mmol/L Li₂SO₄ for 48 h. Nuclei were counterstained with DAPI. Bars = 200 μm. **b** EdU detection in AC16 cells treated with control and 2.5 mmol/L Li₂SO₄ for 48 h. Nuclei were counterstained with DAPI. Bars = 200 μm. **c** Cell number analyses of AC16 cells treated with control and 5 mmol/L Li₂ICl for 48 h. The data are represented as mean ± SEM (n = 3). **p < 0.01, ***p < 0.001. **d** Cell number analyses of AC16 cells treated with control and 2.5 mmol/L Li₂SO₄ for 48 h. The data are represented as mean ± SEM (n = 3). **p < 0.01, ***p < 0.001. **e**, **f** Western blot analyses of the protein levels of PCNA in cells treated with control, 5 mmol/L Li₂ICl and 2.5 mmol/L Li₂SO₄ for 48 h. The data are represented as mean ± SEM (n = 3). **p < 0.001

apoptosis; therefore we test the change of Gsk3 β after lithium treatment. We incubated AC16 cells with LiCl or Li₂SO₄ for 48 h and examined the protein levels of Gsk3 β and phosphorylated Gsk3 β (S9) via western blotting. Interestingly, it showed that LiCl or Li₂SO₄ treatment for 48 h elevated pGsk3 β level significantly (Fig. 5e, f). These results suggest that lithium could control human cardiomyocytes through Gsk3 β signaling.

Discussion

The balance between drug efficacy and cardiotoxicity becomes crucial for clinical trials [26]. Here, we demonstrated that a high concentration of lithium, treatment of bipolar affective disorder, could not only suppress the cell viability of human cardiomyocytes,

but also promote cell apoptosis. Adult cardiomyocytes losing proliferation ability, lack efficient repair when they are injured [25]. Therefore, the slight damage to cardiomyocytes could be accumulated year by year. Importantly, the effects of lithium are long-duration accumulation. For instance, lithium-induced nephropathy seems to be a very slowly progressive disease, and the average period from the initiation of lithium to the presence of end-stage kidney disease is a least 20 years [21]. The toxicity of lithium on the central nervous system and renal has been studied extensively. However, the researches about the chronic poisoning of lithium on the heart remain far from enough. Therefore, it is necessary to pay attention to the large-scale clinical

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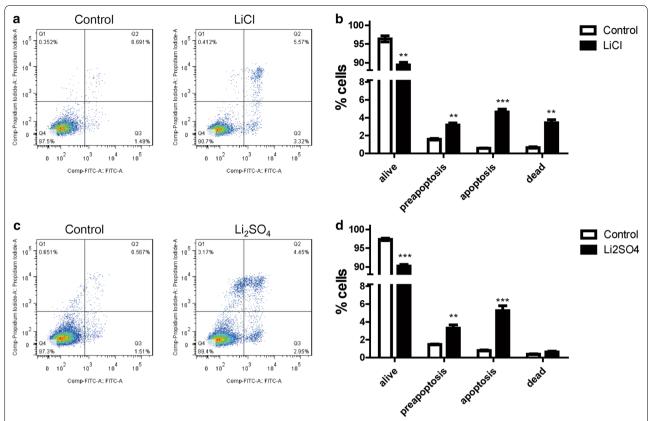


Fig. 4 Lithium triggers the cell apoptosis of human cardiomyocytes. **a** Comparison of AC16 cell apoptosis treated with control or 5 mmol/L LiCl for 48 h via annexin V-FITC/PI apoptosis assay. **b** Statistical analyses of the apoptotic cells in **a**. The data are represented as mean \pm SEM (n = 3).

p < 0.001, *p < 0.001. **c** Comparison of AC16 cell apoptosis treated with control or 2.5 mmol/L Li₂SO₄ for 48 h via annexin V-FITC/PI apoptosis assay. **d** Statistical analyses of the apoptotic cells in **c**. The data are represented as mean \pm SEM (n = 3). **p < 0.001, ***p < 0.001

investigation to detect the damage of lithium to the heart.

It is a fact that the roles of lithium on humans are confusing. It is an important medication option treatment for patients with bipolar disorder [34]. However, the plasma lithium level must be carefully monitored and kept to a narrow range (0.5-1.2 mmol/L) to avoid acute poisoning, such as confusion, seizures and encephalopathy [35]. In addition to acute poisoning, chronic poisoning is the most common side-effect, including reduced urinary concentrating ability, hypothyroidism and weight gain [36]. At the cellular level, lithium reduced cell growth of cancer cells in a dose-dependent manner [37]. Besides, lithium could induce brain iron accumulation and promotes neurodegeneration [38]. These findings, together with our work, suggest the toxicity that we need to pay attention to. In contrast to these defects, low-dose lithium is very important for animals. For instance, it plays an especially important role during early fetal development [39]. In humans, lithium deficiency is associated with increased rates of suicides, homicides and other crimes [39]. Besides, low-dose lithium could protect neurons via inhibiting cell apoptosis [40–42]. Taken together, it is of interest to investigate the roles of low-dose lithium on cardiomyocytes in vitro.

We intake lithium from dietary foods and drinking water, since it is widespread in soils and waters in the earth's crust [7]. Usually, the lithium we intake daily is less than 3 mg, and it might be good for health [39]. Importantly, the lithium level in drinking water, almost negligible at 30 years of age, increases sharply (tap water in Shanghai 0.37 mmol/L), especially for the people living in lithium-rich regions (up to 100 mg/L) [39]. The level of lithium in tap water is much higher than before. Therefore, the lithium we intake increases markedly daily. It is reported that several genes are significantly changing under this level [43]. But the level is much lower than the toxic concentration to humans. Therefore, it could have rare adverse effects on normal adults. However, in lithium-rich regions, the adverse effects need to be determined for specific people such as children who are 2-3 years old. They drink water half as adults drink, while

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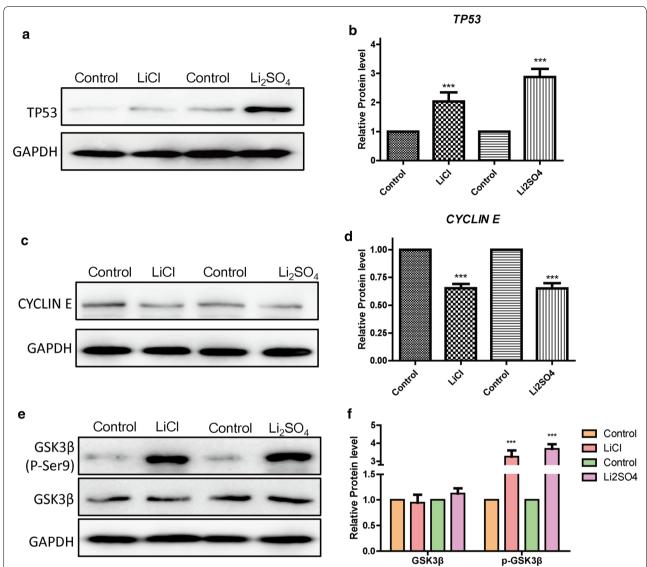


Fig. 5 Lithium controls human cardiomyocytes by Gsk3β signaling. **a** Western blot analyses of the protein levels of TP53 in AC16 cells treated with control, 5 mmol/L LiCl and 2.5 mmol/L Li $_2$ SO $_4$ for 48 h. **b** Statistical analyses of the protein levels of TP53 in **a**. The data are represented as mean \pm SEM (n=3). ***p<0.001. **c** Western blot analyses of the protein levels of Cyclin E in AC16 cells treated with control, 5 mmol/L LiCl and 2.5 mmol/L Li $_2$ SO $_4$ for 48 h. **d** Statistical analyses of the protein levels of Cyclin E in **c**. The data are represented as mean \pm SEM (n=3). ***p<0.001. **e** Western blot analyses of the protein levels of Gsk3β and phosphorylated Gsk3β (P-Ser9) in AC16 cells treated with control, 5 mmol/L LiCl and 2.5 mmol/L Li $_2$ SO $_4$ for 48 h. **f** Statistical analyses of the protein levels of Gsk3β and phosphorylated Gsk3β (P-Ser9) in **e**. The data are represented as mean \pm SEM (n=3). ***p<0.001

their weigh far less than adults. Notably, the development of their nervous system and kidney is immature. Therefore, they are the potential victims of lithium pollution.

Lithium pollution is a new environmental problem that appeared after the 1990s. This study, together with previous researches, confirms that a high concentration of lithium could cause serious injury to the environment and humans, which needs to be paid more attention to [17, 44, 45]. However, at present, the indicators

of environmental monitoring, such as the detection of soil and water, do not include lithium in the Yangtze River Basin. It needs to add lithium as an indicator as soon as possible, especially in the vulnerable areas. Besides, adding the detection of lithium in agricultural irrigation water is particularly important because it is not only closely associated with the crops, but also with our health [44]. Lithium has been listed as a pollutant that leads to environmental harm in irrigation water supplies

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in Australia [10]. The concentration of lithium entering waterways should be less than or equal to 2.5 mg/L [10]. Therefore, it should be added in this area as quickly as possible in the future. In addition to detection, recycling is an effective approach to reduce lithium pollution in the next 20 years [46-48]. To this end, one possible method is to collect taxes on lithium batteries. It could raise the price of lithium batteries, which might improve the benefits of lithium recycling. Therefore, it can not only reduce the waste of resources, but also increase environmental protection. In addition, garbage classification is currently emerging in China, including Shanghai and Beijing. It is a fundamental method for garbage recycling. Thus, the classification and recycling of lithium batteries will significantly increase the efficiency of lithium resources and reduce environmental pollution.

There are some shortages that need to be improved in this study. For example, we found that lithium may affect the activity of cardiomyocytes by regulating the GSK3\beta pathway. Intriguingly, it is reported that the mechanism of lithium changes with different concentrations. At a concentration of 2 mmol/L, it inhibits GSK-3 directly, while at a concentration of 0.8 mmol/L, it regulates protein kinase B (PKB) [49]. Besides, it is reported that lithium could promote protein-coding RNAs that interacted with Alzheimer's disease-associated genes [50]. Furthermore, lithium might interact with adenosine triphosphate (ATP) and form a bimetallic (Mg·Li) ATP complex [51]. Therefore, the mechanism of lithium on cardiomyocytes might be multifaceted. On the other hand, since the water flow in the Yangzi River is constantly changing in the four seasons, the concentration of lithium is changing as well. Therefore, more work will be required to detect it all through the year in the future. Besides, we found that 5 mmol/L lithium inhibited cell viability and promoted cell apoptosis significantly. However, this concentration is higher than the therapeutic concentration (<1.5 mmol/L). Though there is no significant effect at 1 mmol/L, it is of interest to investigate the effects for a long time and further clinical investigation might be valuable.

Conclusions

In summary, here we found that the enormous consumption of lithium batteries could be a serious threat to the environment of the Yangtze River Basin. At the same time, we found that certain concentrations of lithium will cause damage to human cardiomyocytes. It can not only inhibit the activity of cardiomyocytes, but also promote cardiomyocyte apoptosis. Therefore, it is essential to detect lithium pollution in the environment (such as soil and water sources) and effectively recycle lithium batteries in the future.

Abbreviations

EdU: 5-Ethynyl-2'-deoxyuridine; CCK-8: Cell Counting Kit-8; PBS: Phosphate buffer saline; PCNA: Proliferating cell nuclear antigen; TP53: Tumor protein p53; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase; DNA: Deoxyribonucleic acid.

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Not applicable.

Authors' contributions

JL conceived and designed the research; JS and XL performed and designed experiments, analyzed results and wrote the manuscript; XS and WW collected clinical data and performed the experiments; HZ and XW performed the experiments and analyzed the data. All authors read and approved the final manuscript.

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Availability of data and materials

The data supporting the conclusions of this article are included within the article and its additional file.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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